The Pharmaceutical Applications of CRISPR-Cas9 and Its Development Trends

Yu Sang^{1,*}

¹University of Bristol, Bristol, United Kingdom

*Corresponding author: ew21018@ bristol.ac.uk

Abstract:

Originally derived from the immune system of bacteria, CRISPR-Cas9 technology has rapidly developed into a revolutionary gene editing tool that is widely used in molecular biology, drug development, and gene therapy. This technology not only provides a new treatment approach for genetic diseases such as sickle cell anemia and Duchenne muscular dystrophy, but also brings breakthrough progress in cancer treatment by enhancing chimeric antigen receptor T cell (CAR-T) therapy. CRISPR-Cas9 has also shown great potential in the drug discovery stage, especially in gene screening, development of disease models, and drug resistance research. However, with the rapid development of CRISPR technology, the scientific community still needs to solve challenges such as potential off-target effects in gene editing, inefficient delivery systems, and long-term safety. In addition, the ethical issues of gene editing have also sparked widespread discussion, especially how to ensure fair access when treating genetic diseases. Despite many challenges, CRISPR-Cas9 technology still shows great application prospects in the future of drug development and gene therapy.

Keywords: CRISPR-Cas9; pharmaceutical applications; chimeric antigen receptor T cell.

1. Introduction

Originally derived from the bacterial immune system (in which it typically works to edit parts of the genome to protect against viral infections), it has very quickly become one of the most versatile and efficient techniques to engineer genomes, and revolutionised molecular biology, biotechnology and medicine. Since its first application as an engine of gene editing in 2012, it has become a revolutionizing technology, from basic research to efficient development of novel therapeutic strategies for some of our most challenging genetic disorders. [1]

The pharmaceutical industry specifically has applied the Cas9 system to the drug discovery and development stages thanks to its ability to create better disease models and new therapeutical approaches. For instance, by using CRISPR-Cas9, researches have developed novel gene therapies that edit the specific disease-causing mutations in sickle cell anaemia, muscular dystrophy, cystic fibrosis and other genetic disorders. Other therapeutical way of using CRIS-PR-Cas9 is cancer treatments, particularly the use of engineered immune cells such as chimeric antigen receptor T (CAR-T) cells to treat cancers more effectively. The rapid development of CRISPR-Cas9 has also accelerated drug discovery, enabling the creation of more accurate disease models, high-throughput genetic screens and the discovery of new drug targets. In addition, the advancement of CRISPR-Cas9 technology has also been explored to tackle antimicrobial resistance by adding a 'kill switch' (targeted resistance genes in the pathogenic bacteria for deactivation) to resistant bacteria.

Despite many promising ways of applications, CRIS-PR-Cas9 drugs are an emerging class of agents, and would require quite a bit more optimisation – not to mention some difficult conceptual and ethical questions about genetic editing, off-target effects, and effective vector delivery– before they can be used in other fields.

With the high pace of innovation and the enormous potential of CRISPR-Cas9 to improve human health, this review attempts to take a deeper look into what exactly is happening as CRISPR-Cas9 makes its entrance into the pharmaceutical field. This article focuses on the current progress of CRISPR-Cas9 in the pharmaceutical industry, like in the field of gene therapy, introducing its use in cancer as a therapy, and its possible applications in drug discovery and development, to name a few topics. This paper will discuss the future concerns, and subsequently how to address those drawbacks to help CRISPR-Cas9 attain its full potential and achieve what people hope in the field of gene therapeutics.

2. CRISPR-Cas9 in Gene Therapy

Using this cutting-edge tool, CRISPR-Cas9 will enable researchers to edit regions of the genome at very small scale. Originally based on bacterial immune systems, CRISPR has had tremendous success treating genetic disorders such as sickle cell disease (SCD), which is an autosomal recessive blood disorder in millions of people worldwide. The SCD is caused by a mutation that produces dysfunctional blood and can lead to conditions such as pain attacks and organ failure. When CRISPR fixes this mutation, researchers expect it to lead to better drugs or perhaps even a cure for the disease.

One of the first important discoveries was Frangoul et al., who applied CRISPR-Cas9 to the BCL11A gene, which encodes a regulator of haemoglobin production [2]. In general, once it has been initiated, BCL11A represses foetal hemoglobin (HbF). However, by editing BCL11A in SCD patients, researchers successfully reactivated HbF

production. Elevated HbF levels have been shown to reduce the frequency of vaso-occlusive crises, improving patient outcomes. This therapy allows the body's natural processes to manage SCD symptoms more effectively.In addition to BCL11A-based therapies, other approaches are being explored. One example involving directly correcting the β -globin mutation in the HBB gene, which could provide a more direct solution to the genetic defect that causes SCD [3]. Although based on current research, BC-L11A-based strategy is currently more advanced and may offer a quicker path to clinical use, HBB gene correction has the potential for a more complete cure.

Despite initial successes, the long-term safety and efficacy of gene therapies remains uncertain. Esrick et al. provided important information about the efficacy of gene therapy treatment in SCD and demonstrated that their treatment could rapidly reduce symptoms of disease. [4]. However, many gene therapy trials focus on short-to-medium-term results (e.g., 6-18 months). While these studies often show significant improvements in patient outcomes, chronic conditions like SCD demand that treatments remain effective for decades. Continued monitoring will determine whether edited cells persist in the body and maintain their therapeutic effects. Moreover, the possibility of late-onset effects, such as immune reaction arising from unintended gene modifications, means that patient monitoring must extend beyond the early success phase. Thus, more comprehensive data is necessary to demonstrate CRISPR's viability as a long-term treatment for chronic genetic disorders over the period.

Gene therapy does have a potential problem with the delivery system. Recent ex vivo techniques are effective but complex, expensive, and invasive, limiting their use to a wider population [5]. Probably a better way could be the treatment process by employing in-vivo methods where genetic modifications are performed directly on the individual. Despite the potential, in situ protein therapies face issues, particularly in reaching precise targeting of hematopoietic stem cells(HSC) and providing high editing performance. The long-term success of the treatment depends on the ability to target HSC without affecting other cells, as off-target effects could lead to complications like immune reactions or inappropriate gene modifications [5]. Important ethical debates have also been sparked by the rapid development of CRISPR systems for the treatment of genetic disorders. Ormond et al. highlighted key ethical considerations about ensuring everyone has equal access to these life-changing treatments [6]. This is especially true for conditions such as sickle cell disease (SCD), which affects millions of people worldwide, particularly in areas with limited health services, such as Africa. SCD is less prevalent in these areas, so people who need it should

ISSN 2959-409X

be able to get it thanks to whatever advances CRISPR technology produces. Beyond this, there are some general moral concerns, such as whether to treat illnesses while even modifying additional human characteristics, which complicate ethical debates regarding gene editing's limitations.

Also, Brendel et al. have explored combinatorial strategies, such as integrating DNA addition with genome editing, which may increase healing effects by addressing various aspects of the disease together [5]. For instance, a combination of increasing the fetal hemoglobin (HbF) through gene addition and correcting the beta-globin gene mutation through genome editing could improve the disease's control. This might increase blood oxygenation, lessen terrible crises, and improve patient outcomes nevertheless. However, combining these approaches creates new technological complexity as It might be more difficult to get the proper cells, especially in vivo. Safety is another consideration. Combining multiple genetic interventions may increase the chance of off-target effects or unintended consequences, such as immune reactions. Therefore, further research should demonstrate that these combinatorial approaches can be safe, efficient, and scalable for widespread use.

Despite CRISPR-Cas9's early success and gene therapy, there are still obstacles. Perhaps the most important one is getting these therapies widely, especially where the health system is weak. Most developing countries are not wellequipped to provide medical treatments like gene therapy for common genetic disorders like sickle cell disease (SCD), as it requires advanced medical facilities to identify, treat, and observe. Another issue is also the medical treatment's long-term safety. Late-onset negative effects, particularly cancer, must be carefully considered because gene editing can permanently alter the genome. Another issue is the CRISPR-Cas9 efficiency. A significant factor is also the CRISPR-Cas9 technology's precision. Off-target effects, in which other elements of the genome are falsely edited by the gene-editing tool, can lead to unanticipated issues. Coming research should aim to reduce the off-target effects.

Gene therapy for Duchenne Muscular Dystrophy (DMD) could be another potential target. DMD arises because of mutations in the gene dystrophin, one of the biggest genes in the human genome. Because the dystrophin gene is so large, gene replacement has proven difficult However, CRISPR has the potential to identify and correct gene variants to solve this issue.

A new form of "exon skipping" using CRISPR-Cas9 addresses this length requirement. To create a compressed but practical protein, which in this situation is dystrophin, the method involves removing particularly altered exons from a gene. Long and others. demonstrated successful dystrophin restoration and improved muscle function in mouse models [7]. This work laid the foundation for subsequent studies, leading to the breakthrough results reported by Amoasii et al. in dogs. Since dogs share more similarities with humans than smaller animals like mice, this was a crucial step toward applying the treatment in humans. [8].

Researchers continue to refine the method by exploring multiplex gene editing strategies. Ousterout et cetera. demonstrated the ability to target multiple exons at once, potentially addressing a wider range of DMD-causing mutations [9]. The ability of CRISPR-Cas9 to target multiple genes at once shows how flexible this tool can be in treating complex genetic disorders. Other strategies, like increasing the production of utrophin—a protein similar to dystrophin—are also being studied as alternative to directly fixing dystrophin. These different approaches suggest that CRISPR-Cas9 could help target through different molecular pathways.

As the study shift towards clinical applications, safety considerations are the primary concern. Nelson et al. (2019) provided encouraging report that CRISPR-mediated exon skipping shows minimal off-target effects [10]. These findings are crucial for applying this technology towards human trials as the off-target effects are one of the major risks of CRISPR-Cas9 technology, as reviewed comprehensively by Dever et al. [11].

For DMD, the efficacy of in vivo delivery to muscle remains a major challenge. Virus vector-based delivery systems currently have a payload limit, which can only carry a limited amount of genetic material and could be immunogenic. As a result, there is a need for the development of new delivery approaches including lipid nanoparticles and cell-damaging peptides. Additionally, it might not be possible to completely reprogram the entire proteins when producing shorter dystrophin enzymes, despite their use as a therapeutic tool. This requirement may affect the performance of exon-skipping strategies, particularly those that address non-muscle DMD symptoms like cognitive impairment brought on by some dystrophin isoforms.

Another challenge in treating Duchenne Muscular Dystrophy (DMD) with CRISPR-Cas9 is the variety of mutations that cause the disease. A single CRISPR approach may not be effective for everyone because different variants can affect various regions of the dystrophin protein. This means that personalized approaches need to be developed for each person, which could make the diagnostic process more complicated and slower, especially when it comes to getting regulatory authorization for these personalized treatments.

In conclusion, while CRISPR-Cas9 based care for genetic

disorders shows promising results in the clinic, the whole scale implementation in the clinic is difficult. Experts will focus on working out how to get procedures to work more efficiently with cells, fine tune gene editing and make sure that solutions stay in place for a long time and safely and effectively. Although the technology for CRISPR is still being investigated, it show promise for those suffering from these illnesses, and other natural conditions.

3. CRISPR-Cas9 in Cancer Treatment: Revolutionary Potential and Regulatory Challenges

Combining CRISPR-Cas9 technology with cancer immunotherapy (especially with increasing Chimeric Antigen Receptor T-cell therapy, or CAR-T) is a breakthrough in cancer treatment. Using this new technique, scientists will be able to use CRISPR to provide specific gene modifications to to destroy cancer cells. One of the first clinical trials at the University of Pennsylvania, conducted by Dr. Carl June, as published by Stadtmauer et al. (2020), have shown that CRISPR-Cas9 editing of T cells is feasible and even harmless at first [12]. In this trial, three genes-TRAC, TRBC, and PDCD1-were edited to enhance T cell function. By removing the endogenous T-cell receptor and knocking out PD-1, the edited T cells were more resistant to cancer's immune evasion mechanisms. This achievement has suggested new possibilities for CRIS-PR-based cancer therapies.

Based on that, scientists have looked at a variety of strategies to enhance CAR-T therapy using CRISPR. Rupp et al. presented the possibility of generating allogeneic (or "off the shelf") CAR-T cells, which would dramatically shorten the time and cost to manufacture the therapy, and make it more accessible [13]. This overcomes a major issue of existing CAR-T therapies, which are targeted and therefore costly and time-consuming to develop.

While CAR-T therapy has been highly effective for blood cancers, it has faced challenges in treating solid tumors. Choi et al. used CRISPR-Cas9 to engineer CAR-T cells that can better penetrate and survive within the tough environment of solid tumors [14]. Their preclinical experiments in human glioblastoma, one of the most aggressive brain cancers, showed promising results and point towards novel therapies for solid cancers.

But in addition to CAR-T cells, CRISPR can be harnessed to refine other T cell cancer treatments. Li et al. have shown that CRISPR can be harnessed to remove the natural T cell receptor from tumour-infiltrating lymphocytes, thereby making them more selective against cancer [15]. This approach could broaden the impact of cell therapies to treat a wider range of cancers.

Researchers are also combining CRISPR with other cancer treatments to enhance effectiveness. Liu and others found out that CRISPR-engineered CAR-T cells had a stronger impact in animal models when used alongside checkpoint inhibitors [16]. This combined approach could help overcome resistance to treatment and lead to better outcomes.

However, CRISPR-based therapies still face challenges. One major issue is the possibility of off-target effects, where CRISPR modifying the DNA unintentionally. Research such as Nobles et al. have applied advanced sequencing tools to identify these off-target edits in CRIS-PR-modified T cells, providing vital information to enhance the safety and efficacy of these treatments [17].

Additionally, the fast development of CRISPR-edited cell therapies presents unique regulatory challenges. Sherkow and others discussed how governmental agencies, including the FDA, are crafting fresh guidelines to evaluate these therapies [18]. Gene editing represents a relatively new frontier in medicine, and the implications of modifying the human genome—whether somatically or germline—require careful consideration. The evolution of CRISPR-based cancer procedures may have a significant impact on their future.

In conclusion, CRISPR-Cas9 is altering the experience of cancer care, not least as a result of its association with CAR-T body treatment, which offers hope for difficult-to-treat cancers. However, making the technology work successfully, ensuring safety, working around regulatory limitations, and dealing with social issues may prove important.

4. CRISPR-Cas9 for Drug Discovery and Development

In some aspects CRISPR-Cas9 is a direct therapeutic, and its impact on drug discovery and development is profound. Its most popular application is its use in high throughput genetic screens. Screening these effectively lets these scientists delete a single gene at a time and examine it for the effect it has on cellular properties, with the hope of gaining insight into how these work, and more importantly, what makes them a target for drugs. Cancer has proven to be especially well suited for this approach. For example, researchers at the Broad Institute use CRISPR-Cas9 screens to figure out which genes are deleted into cancer cells to make them susceptible to certain drugs. Synthetic lethality is helping to revolutionize cancer treatment and is on the verge of creating fascinating strategies to overcome cancerous drug resistance. However, we have to

ISSN 2959-409X

understand that even these screens with so much promise for novel drug targets aren't 100 % effective. However, using in-vitro methods fails to represent the complexity of the human body. Next, the single gene approach overlooks other important genetic factors as well as multi gene interactions.

CRISPR Cas9 also allows researchers to develop more fine tuned disease versions in animals, so they can better study the effects. We can now understand how conditions develop, and evaluate possible treatments more easily. Additionally, this technology It helps to see how genes work and how medicines affect them, advancing substance. discovery's initial stages. This would enable it shorten the time and the cost to the clients while bringing the new medication.

Yet, along with quick development, there are moral concerns. For example, increased animal testing could be the result of (for example) raising personalized disease models in animals like dogs. Therefore it is important to find the right balance between modernizing technology and attending to the dog. Second, CRISPR based drug discovery could help us understand the long term effects of some of these biological changes. For these reasons, they need to be monitored for safety globally and followed over the long term.

5. Conclusion

CRISPR-Cas9 technology is changing the landscape of genetic disease and cancer treatment, and its application in gene therapy, cancer immunotherapy, and drug development has achieved remarkable results. However, the full realization of the clinical application of this technology still faces technical bottlenecks such as off-target effects, long-term safety, and delivery systems. In addition, ethical and regulatory issues also need to be further explored to ensure the safety and fairness of the technology. With the continuous efforts of researchers, CRISPR-Cas9 is expected to play a greater role in improving human health and conquering difficult diseases in the future, and promote the development of precision medicine and gene therapy.

References

[1] Doudna, J. A., Charpentier, E. The new frontier of genome engineering with CRISPR-Cas9. Science, 2014, 346: 1258096.

[2] Frangoul, H., Altshuler, D., Cappellini, M. D., Chen, Y.-S., Domm, J., Eustace, B. K., Foell, J., de la Fuente, J., Grupp, S., Handgretinger, R., Ho, T. W., Kattamis, A., Kernytsky, A., Lekstrom-Himes, J., Li, A. M., Locatelli, F., Mapara, M. Y., de Montalembert, M., Rondelli, D., Sharma, A., Sheth, S., Soni, S., Steinberg, M. H., Wall, D., Yen, A., Corbacioglu, S. CRISPR- Cas9 gene editing for sickle cell disease and β -thalassemia. New England Journal of Medicine, 2021, 384: 252-260.

[3] Demirci, S., Uchida, N., Tisdale, J. F. Gene therapy for sickle cell disease: An update. Cytotherapy, 2018, 20: 899-910.

[4] Esrick, E. B., Lehmann, L. E., Biffi, A., Achebe, M., Brendel, C., Ciuculescu, M. F., Daley, H., MacKinnon, B., Morris, E., Federico, A., Abriss, D., Boardman, K., Khelladi, R., Shaw, K., Negre, H., Negre, O., Nikiforow, S., Ritz, J., Pai, S.-Y., London, W. B., Dansereau, C., Heeney, M. M., Armant, M., Manis, J. P., Williams, D. A. Post-transcriptional genetic silencing of BCL11A to treat sickle cell disease. New England Journal of Medicine, 2021, 384: 205-215.

[5] Brendel, C., Guda, S., Renella, R., Bauer, D. E., Canver, M. C., Kim, Y. J., Heeney, M. M., Klatt, D., Fogel, J., Milsom, M. D., Orkin, S. H., Gregory, R. I., Williams, D. A. Lineage-specific BCL11A knockdown circumvents toxicities and reverses sickle phenotype. J Clin Invest, 2016, 126: 3868-3878.

[6] Ormond, K. E., Mortlock, D. P., Scholes, D. T., Bombard, Y., Brody, L. C., Faucett, W. A., Garrison, N. A., Hercher, L., Isasi, R., Middleton, A., Musunuru, K., Shriner, D., Virani, A., Young, C. E. Human germline genome editing. Am J Hum Genet, 2017, 101: 167-176.

[7] Long, C., Amoasii, L., Mireault, A. A., McAnally, J. R., Li, H., Sanchez-Ortiz, E., Bhattacharyya, S., Shelton, J. M., Bassel-Duby, R., Olson, E. N. Postnatal genome editing partially restores dystrophin expression in a mouse model of muscular dystrophy. Science, 2016, 351: 400-403.

[8] Amoasii, L., Hildyard, J. C. W., Li, H., Sanchez-Ortiz, E., Mireault, A., Caballero, D., Harron, R., Stathopoulou, T.-R., Massey, C., Shelton, J. M., Bassel-Duby, R., Piercy, R. J., Olson, E. N. Gene editing restores dystrophin expression in a canine model of Duchenne muscular dystrophy. Science, 2018, 362: 86-91.

[9] Ousterout, D. G., Kabadi, A. M., Thakore, P. I., Majoros, W. H., Reddy, T. E., Gersbach, C. A. Multiplex CRISPR/Cas9-based genome editing for correction of dystrophin mutations that cause Duchenne muscular dystrophy. Nature Communications, 2015, 6: 6244.

[10] Nelson, C. E., Wu, Y., Gemberling, M. P., Oliver, M. L., Waller, M. A., Bohning, J. D., Robinson-Hamm, J. N., Bulaklak, K., Castellanos Rivera, R. M., Collier, J. H., Asokan, A., Gersbach, C. A. Long-term evaluation of AAV-CRISPR genome editing for Duchenne muscular dystrophy. Nature Medicine, 2019, 25: 427-432.

[11] Dever, D. P., Scharenberg, S. G., Camarena, J., Kildebeck, E. J., Clark, J. T., Martin, R. M., Bak, R. O., Tang, Y., Dohse, M., Birgmeier, J. A., Jagadeesh, K. A., Bejerano, G., Tsukamoto, A., Gomez-Ospina, N., Uchida, N., Porteus, M. H. CRISPR/Cas9 genome engineering in engraftable human brain-derived neural stem cells. iScience, 2019, 15: 524-535.

[12] Stadtmauer, E. A., Fraietta, J. A., Davis, M. M., Cohen, A. D., Weber, K. L., Lancaster, E., Mangan, P. A., Kulikovskaya, I.,

Gupta, M., Chen, F., Tian, L., Gonzalez, V. E., Xu, J., Jung, I. Y., Melenhorst, J. J., Plesa, G., Shea, J., Matlawski, T., Cervini, A., Gaymon, A. L., Desjardins, S., Lamontagne, A., Salas-Mckee, J., Fesnak, A., Siegel, D. L., Levine, B. L., Jadlowsky, J. K., Young, R. M., Chew, A., Hwang, W. T., Hexner, E. O., Carreno, B. M., Nobles, C. L., Bushman, F. D., Parker, K. R., Qi, Y., Satpathy, A. T., Chang, H. Y., Zhao, Y., Lacey, S. F., June, C. H. CRISPRengineered T cells in patients with refractory cancer. Science, 2020, 367: 73-80.

[13] Rupp, L. J., Schumann, K., Roybal, K. T., Gate, R. E., Ye, C. J., Lim, W. A., Marson, A. CRISPR/Cas9-mediated PD-1 disruption enhances anti-tumor efficacy of human chimeric antigen receptor T cells. Scientific Reports, 2017, 7: 737.

[14] Choi, B. D., Yu, X., Castano, A. P., Darr, H., Henderson, D. B., Bouffard, A. A., Larson, R. C., Scarfò, I., Bailey, S. R., Gerhard, G. M., Frigault, M. J., Leick, M. B., Schmidts, A., Sagert, J. G., Curry, W. T., Carter, B. S., Maus, M. V. CRISPR-Cas9 disruption of PD-1 enhances activity of universal EGFRvIII

CAR T cells in a preclinical model of human glioblastoma. Journal for ImmunoTherapy of Cancer, 2019, 7: 304.

[15] Li, D., Li, X., Zhou, W.-L., Huang, Y., Liang, X., Jiang, L., Yang, X., Sun, J., Li, Z., Han, W.-D., Wang, W. Genetically engineered T cells for cancer immunotherapy. Signal Transduction and Targeted Therapy, 2019, 4: 35.

[16] Liu, X., Zhang, Y., Cheng, C., Cheng, A. W., Zhang, X., Li, N., Xia, C., Wei, X., Liu, X., Wang, H. CRISPR-Cas9-mediated multiplex gene editing in CAR-T cells. Cell Research, 2017, 27: 154-157.

[17] Nobles, C. L., Sherrill-Mix, S., Everett, J. K., Reddy, S., Fraietta, J. A., Porter, D. L., Frey, N., Gill, S. I., Grupp, S. A., Maude, S. L., Siegel, D. L., Levine, B. L., June, C. H., Lacey, S. F., Melenhorst, J. J., Bushman, F. D. CD19-targeting CAR T cell immunotherapy outcomes correlate with genomic modification by vector integration. J Clin Invest, 2020, 130: 673-685.

[18] Sherkow, J. S., Zettler, P. J., Greely, H. T. Is it 'gene therapy'? Journal of Law and the Biosciences, 2018, 5: 786-793.